

HIGHLY UNSATURATED AMIDES FROM *SALMEA SCANDENS*

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Abstract—The aerial parts of *Salmea scandens* gave, in addition to three known amides, three new ones. The chemotaxonomy of this genus is discussed briefly.

The small tropical American genus *Salmea* has not been studied chemically. We have therefore investigated *Salmea scandens* (L.) DC. The aerial parts contain germacrene D, squalene, nerolidol, spathulenol and the isobutylamide 1, the phenylethylamides 2 and 3, which have been isolated from a *Spilanthes* species [1], as well as three further ones, the epoxide 4, the enzyme 5, and the aldehyde 6. The structure of 5, molecular formula $C_{18}H_{21}NO$, followed from the 1H NMR spectrum (see Experimental), which clearly showed the presence of a phenylethylamide residue. Spin decoupling allowed the assignment of all signals. The couplings of H-2 and H-3 indicate a Z-configuration of the conjugated double bond. The presence of two methylene groups followed by a ynone chromophore also followed from the 1H NMR signals as H-5 was a narrowly split triplet which collapsed to a clear triplet by irradiation of the broad double-quartet at δ 5.42 (H-8). As this signal showed an 11 Hz coupling this double bond also had a Z-configuration.

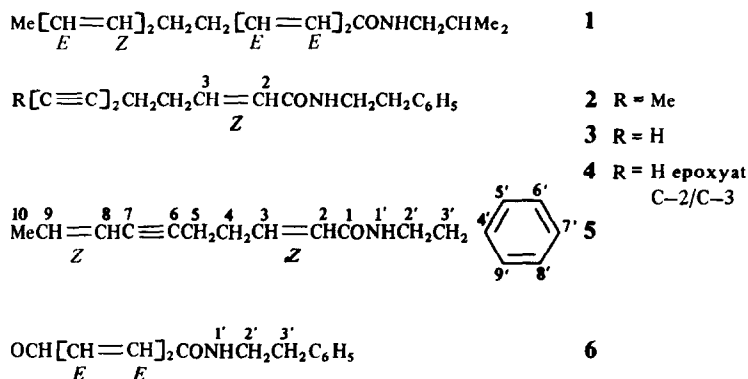
The 1H NMR spectrum of 4 (see Experimental), which gave no molecular ion by the EI technique, but a clear $[M+1]^+$ ion by chemical ionization, showed that an epoxide was present. Thus H-2 was a doublet at δ 3.49 ($J = 5$ Hz) which was coupled with a double-triplet at δ 3.23. The presence of a diyne end group followed from the mass spectrum and the typical long range coupling of the acetylenic proton.

The 1H NMR spectrum of 6 (see Experimental) indicated by the presence of a low-field doublet at δ 9.64 that an aldehyde was present. Again only by chemical ionization was a $[M+1]^+$ peak obtained. All 1H NMR signals were assigned by spin decoupling. Most likely 6 was formed by oxidative degradation of an amide similar to 1 with a phenylethylamide residue.

The chemistry of *Salmea* seems to be very similar to *Spilanthes* and *Acmella*, where highly unsaturated amides are common [1–5]. These compounds are also present in species of *Echinacea* [6] and *Heliopsis* [7]. Bolick [8] has discussed the cladistic relationships of *Salmea* and relates the genus directly to *Spilanthes* and *Acmella*, as do the chemical data. Indeed, several species of *Salmea* previously placed in *Spilanthes* by earlier workers have recently been transferred back to *Salmea* [9]. Robinson [10] comments upon the close relationship of *Salmea* and *Acmella* to *Verbesina* but Bolick reckons the relationship of this complex to be with *Otopappus*, a genus whose chemistry is inadequately known at present.

EXPERIMENTAL

The air-dried aerial parts (460 g, collected in Mexico, voucher Turner and Tapia 15518, TEX) were extracted with MeOH–Et₂O–petrol, 1:1:1, at room temp. and worked up in the usual fashion [11]. CC fractions were as follows: 1 (petrol and



Et₂O-petrol, 1:9), 2 (Et₂O-petrol, 1:1) and 3 (Et₂O and Et₂O-MeOH, 9:1). TLC (petrol) of fraction 1 gave 40 mg germacrene D and 860 mg squalene. TLC of fraction 2 (Et₂O-petrol, 1:1) gave 6 mg nerolidol and 21 mg spathulenol. TLC of 150 mg of fraction 3 (total 21 g) (Et₂O-petrol, 4:1) gave two crude bands (3/1 and 3/2) which were separated further by HPLC (RP 8, MeOH-H₂O, 3:2, ca 100 bar, flow rate 3 ml/min), 3/1 affording 3.8 mg 3 (*R*_f 7.3 min), a mixture (3/1/2) (*R*_f 9.4 min), 5 mg 5 (*R*_f 12.1 min) and 6 mg 1 (*R*_f 20.1 min). TLC (Et₂O-petrol, 4:1, 3 developments) and HPLC (see above) of 3/1/2 gave 2 mg 2 (*R*_f 10.1 min) and 0.6 mg 6 (*R*_f 0.5 min). TLC (CH₂Cl₂-C₆H₆-Et₂O, 1:1:1) of 3/2 gave 8 mg 4 (*R*_f 0.6). Known compounds were identified by comparing the 400 MHz ¹H NMR spectra with those of authentic material.

Nona-6,8-diyne-2,3-epoxy-1-oiic acid phenylethylamide (4). Colourless oil; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3425, 1690, 1520 (CONHR), 3320, 2200 (HC≡C); CIMS (isobutane) *m/z* (rel. int.): 268 [M + 1]⁺ (47), 177 [268 - C₇H₇]⁺ (100); ¹H NMR (CDCl₃, 400 MHz, TMS as internal standard): δ 3.49 (*d*, H-2), 3.23 (*dt*, H-3), 1.41 (*ddt*, H-4), 1.63 (*ddt*, H-4'), 2.35 (*dt*, H-5), 2.05 (*t*, H-9), 3.67 (*dq*, H-2'), 3.49 (*dq*, H-2''), 2.87 (*dt*, H-3'), 2.79 (*dt*, H-3''), 7.18 (*br d*, H-5', H-9'), 7.22 (*t*, H-7'), 7.31 (*t*, H-6', H-8'); [*J* (Hz): 2, 3 = 5, 3, 4 = 8; 4, 4' = 14; 4, 5 = 7; 5, 9 = 1; NH, 2' = 2', 3' = 7; 2', 2'' = 3', 3'' = 14, 5', 6' = 6', 7' = 8].

Deca-2Z,8Z-dien-6-yn-1-oiic acid phenylethylamide (5). Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3470, 1675, 1500 (CONHR), 3025, 1600 (CH=CH); MS *m/z* (rel. int.): 267.162 [M]⁺ (27) (calc. for C₁₈H₂₁NO: 267.162), 252 [M - Me]⁺ (7), 176 [M - C₇H₇]⁺ (8), 147 [M - C₉H₁₂]⁺ (29), 123 [176 - C₄H₅]⁺ (100), 120 [C₉H₁₂]⁺ (63), 119 [C₉H₁₁]⁺ (60), 105 [C₈H₉]⁺ (95), 104 [C₈H₈]⁺ (60), 91 [C₇H₇]⁺ (61), 79 [C₆H₇]⁺ (54), 77 [C₆H₅]⁺ (55); ¹H NMR (CDCl₃, 400 MHz, TMS as internal standard): δ 5.71 (*dt*, H-2), 6.07 (*dt*, H-3), 2.82 (*ddt*, H-4), 2.42 (*dt*, H-5), 5.42 (*dq*, H-8), 5.90 (*dq*, H-9), 1.82 (*dd*, H-10), 3.36 (*t*, H-2'), 2.85 (*t*, H-3'), 7.20 (*br d*, H-5', H-9'), 7.24 (*t*, H-7'), 7.31 (*t*, H-6', H-8'), 5.62

(*br s*, NH); [*J* (Hz): 2, 3 = 8, 9 = 11; 2, 4 = 5, 8 = 2; 3, 4 = 4, 5 = 9, 10 = NH, 2' = 2', 3' = 7; 5', 6' = 6', 7' = 8].

6-Oxo-hexa-2E,4E-dien-1-oiic acid-N-phenylethylamide (6). Colourless oil; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3440, 1680, 1500 (CONHR), 2730, 1690 (CHO); CIMS *m/z* (isobutane) *m/z* (rel. int.): 230 [M + 1]⁺ (100); ¹H NMR (CDCl₃, 400 MHz, TMS as internal standard): δ 6.19 (*d*, H-2), 7.37 (*dd*, H-3), 7.11 (*dd*, H-4), 6.40 (*dd*, H-5), 9.64 (*d*, H-6), 3.65 (*dt*, H-2'), 2.88 (*t*, H-3'), 7.20 (*br d*, H-5', H-9'), 7.25 (*t*, H-7'), 7.33 (*t*, H-6', H-8'); [*J* (Hz): 2, 3 = 4, 5 = 15; 3, 4 = 11; 5, 6 = 7.5; NH, 2' = 2', 3' = 7; 5', 6' = 6', 7' = 8].

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